

# Lawson: a novel reagent for the detection of latent fingerprints on paper surfaces

Renee Jelly,<sup>a</sup> Simon W. Lewis,<sup>\*a</sup> Chris Lennard,<sup>b</sup> Kieran F. Lim<sup>†c</sup> and Joseph Almog<sup>d</sup>

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Lawson (2-hydroxy-1,4-naphthoquinone) reacts with latent fingerprint deposits on paper surfaces to yield purple-brown impressions of ridge details which are also photoluminescent; this compound represents the first in a completely new class of fingerprint detection reagents.

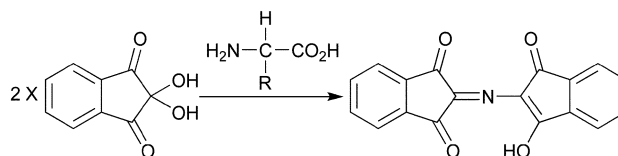
Fingerprints remain the most widely used and reliable means of personal identification and, hence, are extremely important to law enforcement as contact evidence.<sup>1</sup> A key element in the successful recovery of a latent (invisible) fingerprint from a scene or object is detection; to this end, a range of physical and chemical methods are available for the visualisation of such fingerprints.<sup>1,2</sup>

Amino acids are a key component of the secretions that contribute to latent fingerprints<sup>3</sup> and are particularly important when porous surfaces, such as paper, are examined, as they tend to bond to paper fibres and are thus long-lived.<sup>2</sup> Ninhydrin (2,2-dihydroxy-1,3-indanedione) was the first amino acid sensitive reagent for developing fingerprints on porous surfaces and is still the most widely used. Ninhydrin reacts with amino acids to form a dark purple coloured compound known as Ruhemann's purple (Scheme 1).<sup>2</sup>

These treated impressions may be further enhanced by treatment with zinc and cadmium salts to produce complexes that exhibit photoluminescence when cooled to liquid nitrogen temperature (−196 °C) and excited with high intensity light sources.<sup>2</sup>

Since the early 1980s, a range of ninhydrin analogues have been synthesised and tested for their suitability as reagents for the detection of latent fingerprints. Despite the large body of work carried out, only two such reagents have found widespread operational use: 1,8-diazafluoren-9-one (DFO) and 1,2-indanedione<sup>1,2,4</sup> (Fig. 1). More recently, Almog and co-workers have suggested the natural product genipin as an operationally safer and environmentally benign fingerprint reagent exhibiting both colour and photoluminescence.<sup>5,6</sup>

Traditionally genipin has been used as a fabric and skin dye, and this led us to investigate the potential of other natural products, which have been used in this fashion, as fingerprint



Scheme 1 Reaction of ninhydrin with amino acids.

reagents. Henna, a natural product sourced from the leaves of *Lawsonia inermis*, in a similar manner to genipin, has been used as a skin and hair dye for millennia, with reports of its use dating back to 1400 BC.<sup>7</sup> Lawson (2-hydroxy-1,4-naphthoquinone, Fig. 2) is the compound thought to be responsible for the staining properties of henna.<sup>7</sup> This compound was thus selected to be investigated for its ability to develop latent fingerprints on paper surfaces.

Lawson is a naphthoquinone, a group of compounds that are well known for their reactions with amino acids. 1,2-Naphthoquinone-4-sulfonate has been applied to the determination of amino acids through the formation of highly coloured compounds<sup>8–10</sup> and 1,2-naphthoquinone has been reported as yielding purple-brown pigments with cysteine and proteins.<sup>11</sup> Spectroscopic evidence indicates that the reaction of 1,2-naphthoquinone occurs as a result of the primary amine component of amino acids.<sup>11</sup> Intriguingly, a paper describing investigations into the reactivity of the phenyliodonium ylide of 2-hydroxy-1,4-naphthoquinone with amino compounds suggests the formation of indanedione-2-carbox-amido compounds,<sup>12</sup> thus offering an alternative window on the reaction mechanism of 1,2-indanedione.<sup>13</sup> Naphthoquinones therefore represent a class of compounds of significant interest as potential fingerprint detection reagents.

Latent fingerprints were collected on filter paper from a number of different volunteers. Volunteers were requested not to wash their hands immediately before collecting impressions; the fingers were not “charged” with additional secretions in any way. The filter paper strips were then dipped in the lawson reagent solution, air dried and then heated, either in an oven or by direct heat using a laundry press. Initial experiments with lawson in ethanolic solution yielded purple-brown developed marks, which also exhibited strong

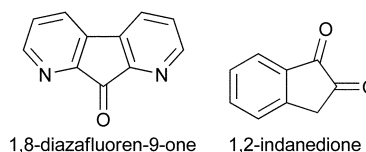


Fig. 1 Amino acid targeting reagents for the development of latent fingerprints.

<sup>a</sup> Department of Applied Chemistry, Curtin University of Technology, GPO Box U1987, Perth, Western Australia 6845, Australia. E-mail: S.Lewis@curtin.edu.au; Fax: +61 8 9266 2300; Tel: +61 8 9266 2484

<sup>b</sup> National Centre for Forensic Studies, Faculty of Applied Science, University of Canberra, Canberra, ACT 2601, Australia

<sup>c</sup> School of Life and Environmental Sciences, Deakin University, Geelong, Victoria 3217, Australia

<sup>d</sup> Casali Institute of Applied Chemistry, The Hebrew University of Jerusalem, Jerusalem 91904, Israel

<sup>†</sup> Lim Pak Kwan (林百君).

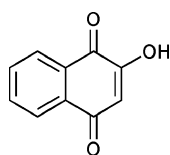


Fig. 2 Chemical structure of lawsone.

photoluminescence when illuminated with a forensic light source (Polilight PL500, Rofin, Australia) at 555 nm and viewed through red goggles (Fig. 3).

The photoluminescence was further investigated using a Cary fluorescence spectrophotometer (Varian, Mulgrave, Australia) with a fibre optic probe attachment (Varian, Mulgrave, Australia), and it was determined that the maximum intensity of luminescence occurred at around 640 nm with excitation at 590 nm. This is operationally significant as photoluminescence emission at longer wavelengths has the potential to improve detectability by avoiding native background luminescence.

It was found that using ethanol as a carrier solvent led to a significant discoloration of the paper substrate, resulting in a degradation of image quality. Ethanol would be unsuitable as a carrier solvent for operational use due to its polarity, which would cause problems with written document evidence due to ink diffusion through the paper. We investigated methyl nonafluoroisobutyl ether (HFE-7100) as the carrier solvent for the working solution; this solvent is used widely for fingerprint reagents such as 1,2-indanedione.<sup>2</sup> In order to successfully dissolve lawsone, a co-solvent of higher polarity was required. Ethyl acetate (20%) was found to be the most successful of the solvents investigated. We also found that petroleum spirit 60–80 °C could substitute for HFE-7100 without a serious impact on the quality of the developed fingerprints.

Preliminary studies established that a concentration of 1 mg lawsone mL<sup>-1</sup> gave the best compromise between contrast and background interference with excess concentrations leading to discoloration of the paper substrate. The formulation used for the remainder of the study was prepared in the following fashion: lawsone (50 mg; Sigma-Aldrich) was dissolved in ethyl acetate (10 mL; Univar) and the resulting solution was subsequently mixed with HFE-7100 (40 mL; 3 M Novec). Latent fingerprint samples on filter paper were dipped in this solution, air dried and then developed with heat. Heating in an oven for 1 h at 140–170 °C produced more uniform development than heating with a laundry press, providing enhancement sufficient for complete visualisation of fingerprint ridge details.

In order to verify that lawsone was reacting with the amino acid content of the latent fingerprints, solutions of amino acids in water (lysine, alanine, glycine, proline; all at 900 µg mL<sup>-1</sup>) were dispensed (10 µL) onto filter paper and allowed to air dry before subsequent lawsone treatment. In common with the developed latent fingerprints, the amino acid spots, with the exception of proline, developed as purple-brown stains which were also photoluminescent. Fluorescence spectra of the developed amino acid spots and a developed latent fingerprint are presented in Fig. 4. It is clear from the profiles that the photoluminescent characteristics of the product of the reaction

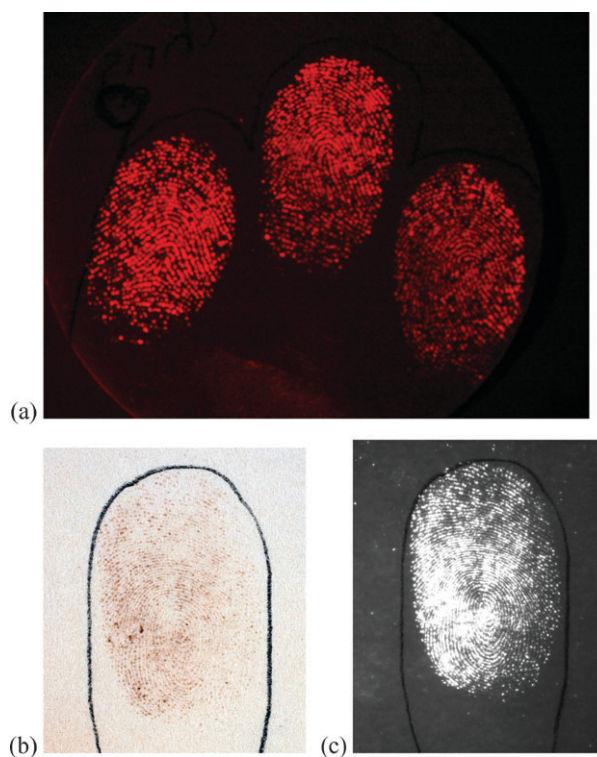


Fig. 3 Lawsone treated latent fingerprints. Images (a) and (b) were taken with a Pentax K10 digital SLR, 50 mm focal length, ISO 100. (a) Photoluminescence mode (excitation with a Polilight PL 500 at 590 nm and viewed through a Wratten NA29 filter, shutter speed 6.0 s, aperture f2.8). (b) Taken under white light (shutter speed 1/125 s, aperture f4). (c) Acquired using a Poliview digital imaging system (Rofin, Australia) with excitation at 590 nm, viewed through a 650 nm interference filter with a 1 s exposure time.

between primary amino acids and lawsone are similar to that in lawsone developed fingerprints, thus confirming that the lawsone reagent is non-specifically targeting primary amino acids in the latent fingerprint deposit. This is a key requirement of latent fingerprint reagents, as the amino acid content of natural secretions is highly variable from one subject to another.<sup>3</sup>

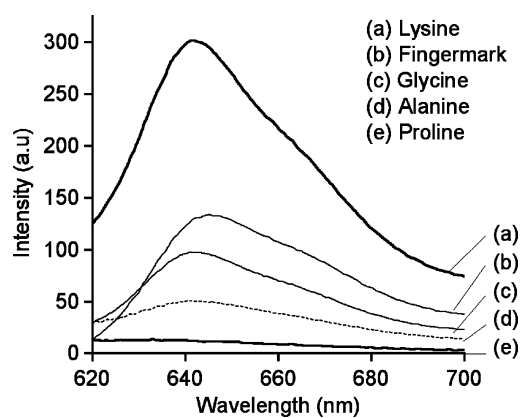
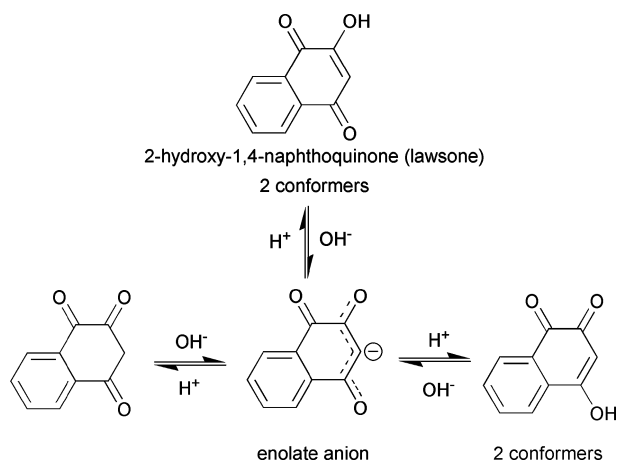
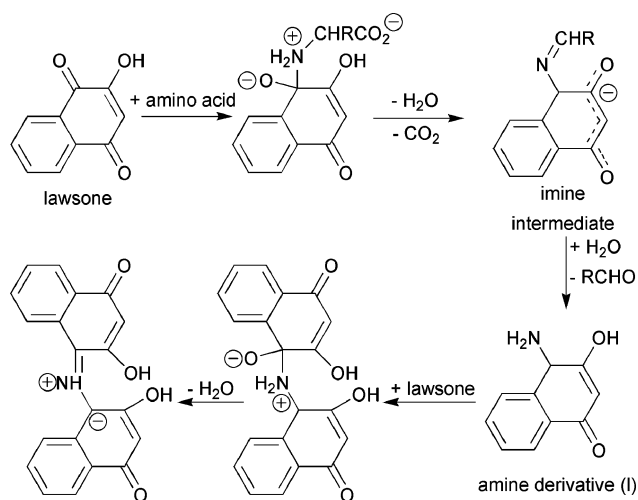


Fig. 4 Fluorescence spectra of a lawsone developed latent fingerprint and of selected amino acids on filter paper (excitation wavelength 590 nm).



Scheme 2 Lawsone and isomers.



Scheme 3 Proposed reaction pathway and product for the reaction of lawsone with primary amino acids.

Lawsone can undergo keto–enol tautomerisation as shown in Scheme 2. Spartan '04<sup>14</sup> *ab initio* calculations using HF/6 31G\* theory indicate that the 1-position of all 6 species/conformers is most likely to react with an incoming nucleophile.

Based on the known reactivity of hydroxyquinones<sup>15</sup> and ninhydrin,<sup>16</sup> we postulate that lawsone undergoes a Strecker degradation<sup>17</sup> at the 1-position to form the amine intermediate (I), as shown in Scheme 3. The Strecker degradation requires a primary amino acid,<sup>17</sup> which would explain why proline does not form the fluorescent product. The amine (I), is a nucleophilic base which can react with a second lawsone molecule to reduce the ketone at the 1'-position<sup>18</sup> to form the final

product, in which the negative charge can be delocalised over the extended  $\pi$ -system.

Further research is required to investigate the reaction mechanism involved and optimise the development conditions for this fingerprint detection reagent. In addition, there is the potential that lawsone could find wider application as a fluorogenic reagent for amino acids; however, additional studies are required to determine optimum reaction conditions in solution and sensitivity *versus* existing reagents.

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## Notes and references

1. J. Almog, in *Encyclopedia of Forensic Sciences*, ed. J. Siegel, P. Saukko and G. Knupfer, Academic Press, San Diego, 2000, vol. 2, pp. 890–900.
2. C. Champod, C. Lennard, P. Margot and M. Stoilovic, *Fingerprints and Other Ridge Skin Impressions*, CRC Press, Boca Raton, 2004.
3. S. Bramble and J. Brennan, in *Encyclopedia of Forensic Sciences*, ed. J. Siegel, P. Saukko and G. Knupfer, Academic Press, San Diego, 2000, vol. 2, pp. 862–869.
4. D. B. Hansen and M. M. Joullié, *Chem. Soc. Rev.*, 2005, **34**, 408–417.
5. J. Almog, Y. Cohen, M. Azoury and T.-R. Hahn, *J. Forensic Sci.*, 2004, **49**, 255–258.
6. G. Levinton-Shamuilov, Y. Cohen, M. Azoury, A. Chaikovskiy and J. Almog, *J. Forensic Sci.*, 2005, **50**, 1367–1371.
7. R. Petkewich, *Chem. Eng. News*, 2006, **84**, 28.
8. E. Frame, J. Russell and A. Wilhelmi, *J. Biol. Chem.*, 1943, **149**, 255–270.
9. N. H. Furman, G. H. Morrison and A. F. Wagner, *Anal. Chem.*, 1950, **22**, 1561–1562.
10. J. Saurina, S. Hernandez-Cassou and R. Tauler, *Anal. Chem.*, 1995, **67**, 3722–3726.
11. J. R. Rees and A. Pirie, *Biochem. J.*, 1967, **102**, 853–863.
12. K. Spagou, E. Malamidou-Xenikaki and S. Spyroudis, *Molecules*, 2005, **10**, 226–237.
13. O. Petrovskaia, B. M. Taylor, D. B. Hauze, P. J. Carroll and M. M. Joullié, *J. Org. Chem.*, 2001, **66**, 7666–7675.
14. *Spartan '04*, available from Wavefunction, Inc., 18401 Von Karman Avenue, Suite 370, Irvine, CA 92612, USA, 2004, <http://www.wavefun.com>.
15. S. Spyroudis, *Molecules*, 2000, **5**, 1291–1330.
16. D. McCaldin, *Chem. Rev.*, 1960, **60**, 39–51.
17. A. Schonberg and R. Moubacher, *Chem. Rev.*, 1952, **50**, 261–277.
18. J. McMurry, *Organic Chemistry*, Brookes/Cole, Pacific Grove (CA), 6th edn, 2004.